

## **cDNA analyses of the MMR genes**

***MLH1, MSH2, MSH6, and PMS2***

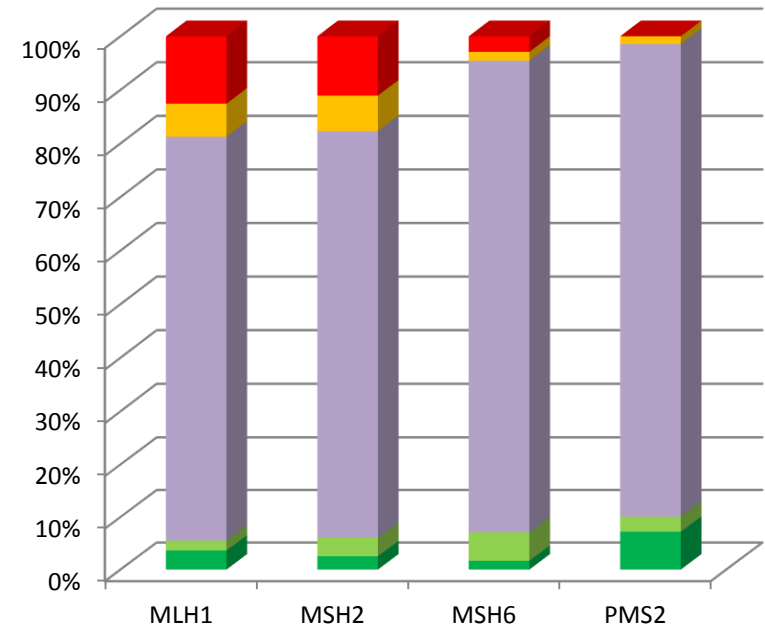
**investigate the effect of VUS upon splicing,  
detect unexpected splicing defects,  
and find allelic losses indicating a germline defect**

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# Pathogenicity assessment by RNA-analysis



- missense variants
- splice site variants
- intronic
- synonymous
- 5' / 3' UTR
- exon duplication
- ins/del of one amino acid



## Prerequisites for RNA based pathogenicity assessment

- Allelic balance
- Normal splicing pattern
- Efficiency of NMD
- Abnormal splicing pattern

⇒ **Cut-off definition by analysing more than 900 cDNAs**

⇒ **Full length cDNA analysis protocol - EMMR working group**

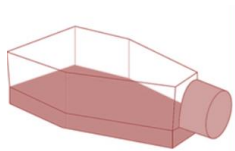
# Isolation of patient's total RNA

## RNA derived from patient blood samples

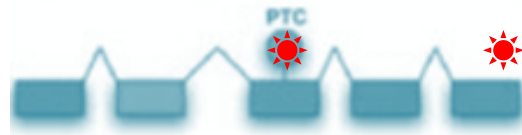
2x heparine



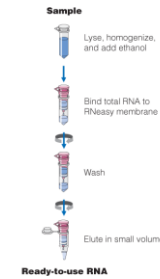
short-term lymphocyte cultures



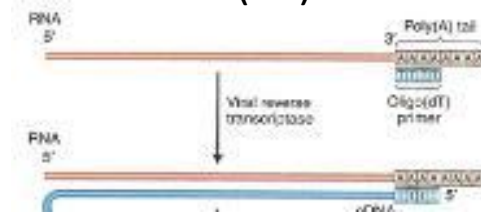
**NMD-inhibition**  
+puromycin  
-puromycin



RNA isolation

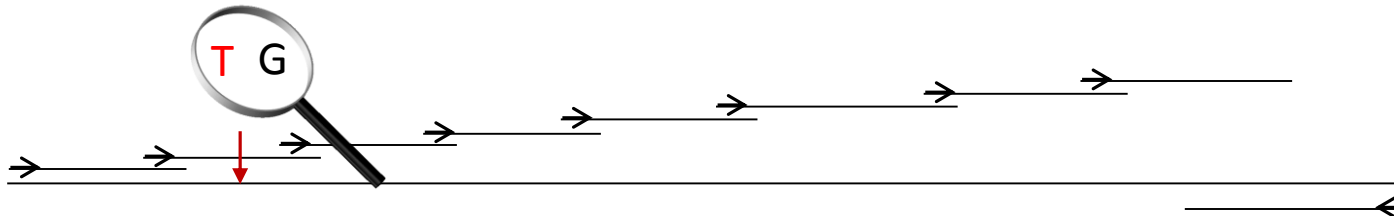


cDNA synthesis  
dT(20)



72-96 h, 37°C **Nonsense-Mediated-mRNA-Decay (NMD)**  
early truncating > 55 nt prior to last exon-junction

48°C, 75-90 min



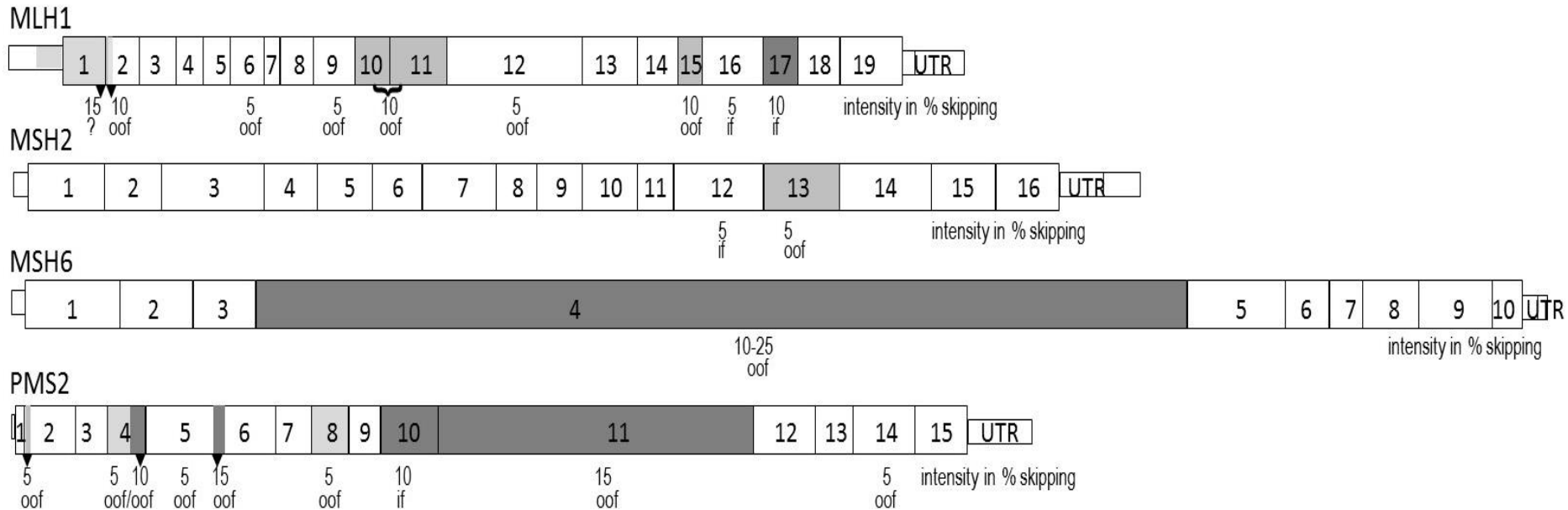
*MLH1* ~ 2.5 kb

*MSH2* ~ 3 kb

*MSH6* ~ 4.2 kb

*PMS2* ~ 2.8 kb

# Alternative splicing in controls



-> alternative splicing in cDNA-P and cDNA+P usually 0-10%

-> gene- and exon-specific exceptions with higher values

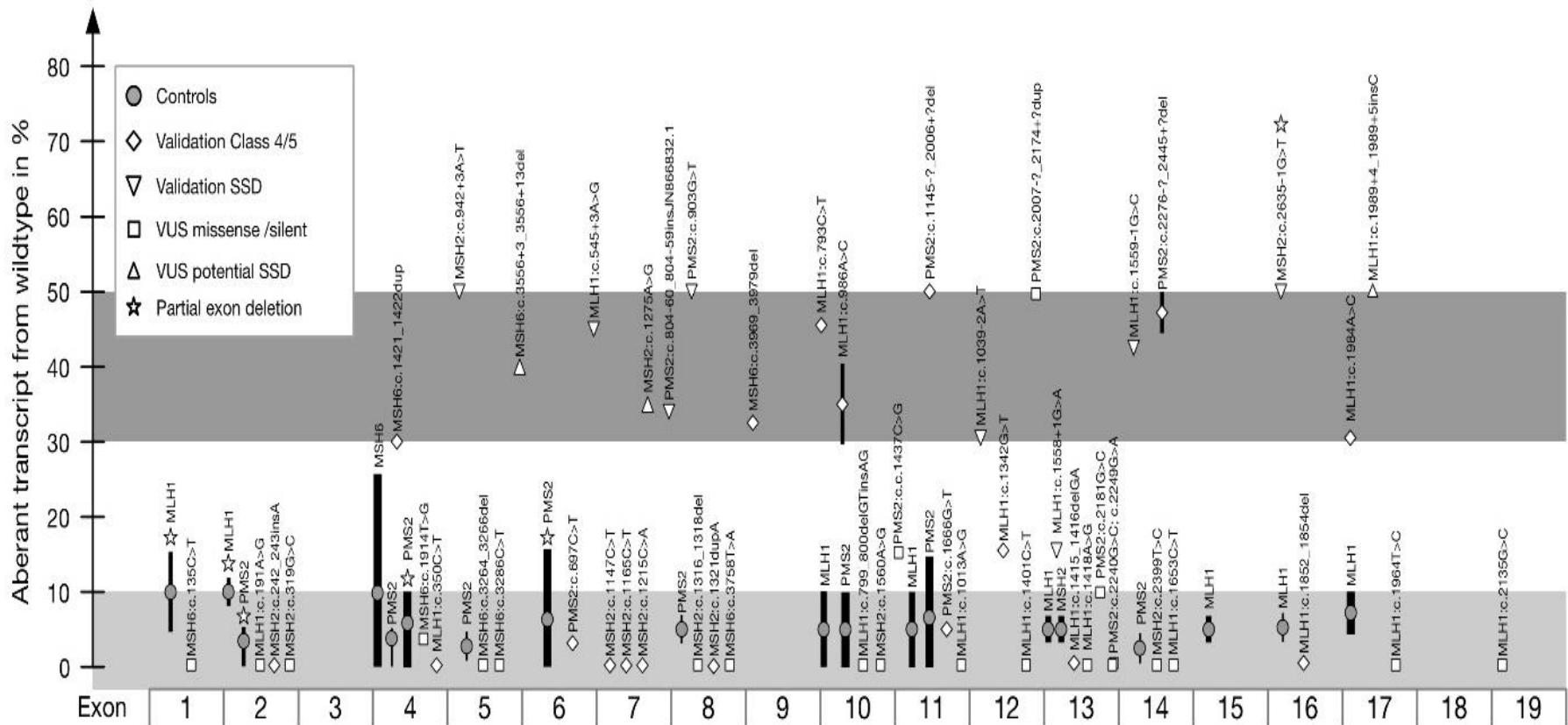
of up to 15% for *MLH1* exon 1q, *PMS2* exon 6p and exon 11,

of up to 25% for *MSH6* exon 4

# Cut off definition

Allelic balance	50% +/- 10%
Allelic loss	< 10%
Normal splicing	up to 10%, some exceptions
Aberrant splicing	above 30%

Universal protocol used within the european mismatch repair consortium





# Testing 5 Class 5 variants

5 variants predicted to affect one amino acid (putative missense/indel), class 4-5:

## ***MLH1* class 4-5 variants**

- c.793C>T p.(Arg265Cys)
- c.986A>C p.(His329Pro)
- c.1984A>C p.(Thr662Pro)
- c.1852\_1854del p.(Lys618del)
- c.350C>T p.(Thr117Met)

## **bioinformatical prediction:**

RNA analysis 5 not predicted as splice defects

protein function 3 predicted deficiency

## **Effect on mRNA splicing**

- c.793C>T p.(Arg265Cys)
- c.986A>C p.(His329Pro)
- c.1984A>C p.(Thr662Pro)
- c.1852\_1854del p.(Lys618del)
- c.350C>T p.(Thr117Met)

## **experimental analysis:**

**splice site defect: 2 complete, 1 partial**

5 MMR deficiency

# Testing 25 class 3 variants



## variant

3 potential SSD  
13 missense variants  
1 exon duplication  
4 synonymous variants  
4 VUS + pathogenic variant

## cDNA result

all splicing-defects  
all splice-neutral, biallelic  
duplication verified  
all splice-neutral, biallelic  
all splice-neutral, biallelic

## re-classification

2x class 5 (complete SSD)  
0, remain class 3 due to unclear  
1x class 4 (funct. protein domain)  
4x class 2  
2x re-classification to class 2

-> re-classification of 36% of VUS

-> splicing defects 12%

# Testing MMR deficient unsolved patients



## 26 „mutation-negative“ patients with unsolved IHC loss in CRC:

MLH1/PMS2 in 16 patients

PMS2 in 5 patients

MSH2/MSH6 in 2 patients

MSH6 in 3 patients

### ➤ 5/26 abnormal cDNA results (19%)

#### cDNA result

*MLH1* NMD-P, PTC+P

*MLH1* NMD-P, SSD oof+P

*PMS2* NMD-P, SSD ins oof+P

*MLH1* allelic loss -P/+P

*MLH1* allelic loss -P/+P

#### genomic cause

*MLH1* truncating variant (overseen)

*MLH1* splice site variant (overseen)

*PMS2* partial intron inclusion

?

?

### ➤ 21/26 normal FLT = no germline deficiency in MMR gene analyzed



# Summary



- **protocol for full-length cDNA analysis for the four MMR transcripts**
  - **definition of cut-off values for allelic representation, normal and aberrant splicing**
  - **testing the system on different variants**
    - missense variants (class 4-5) -> 60% splicing defects**
    - 25 different types of VUS class 3 -> 12% splicing defects, 36% re-classified**
  - **testing the system on 26 MMR deficient, unsolved patients**
    - 19% with abnormal cDNA result = MMR-defect**
    - 81% with normal cDNA results = no MMR-defect in germline**
- ⇒ **Transfer to NGS-bases RNA-analysis by using these data to calibrate the system and educate bio-IT systems**
- ⇒ **Creating a meaningful algorithm for pathogenicity assessment**

# Thanks to all contributors



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MVZ Labor Dr. Fenner und Kollegen, Hamburg

# cDNA analysis using NGS technologies

Next step:

Lab-project

Use the defined cut-offs to validate  
automated, NGS-based cDNA analyses

IT-project

Improve skills of bio-IT



# Classification of MMR gene variants



## InSiGHT guidelines

class	1	2	3	4	5
	non-pathogenic	probably non-pathogenic	VUS uncertain significance	probably pathogenic	pathogenic

Variant Interpretation Committee



- splice site variants
- intronic
- synonymous
- 5`/ 3`UTR
- exon duplication
- predicted missense variants
- ins/del of one amino acid



- mRNA splicing/expression
- ↓
- functional protein assays

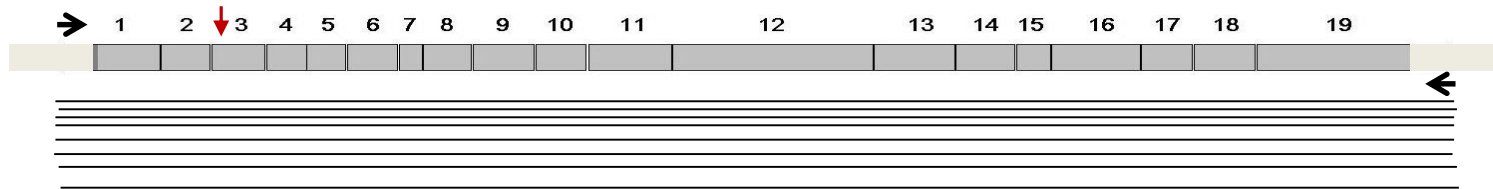
=> RNA based analysis for pathogenicity assessment of sequence variants



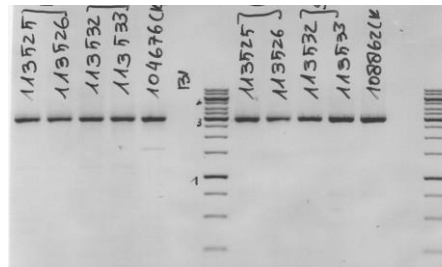
# Full-length transcript amplification and sequencing



- LR-PCR from cDNA: primers in first and last exon



- agarose gel electrophoresis



*MLH1* ~ 2.5 kb

*MSH2* ~ 3 kb

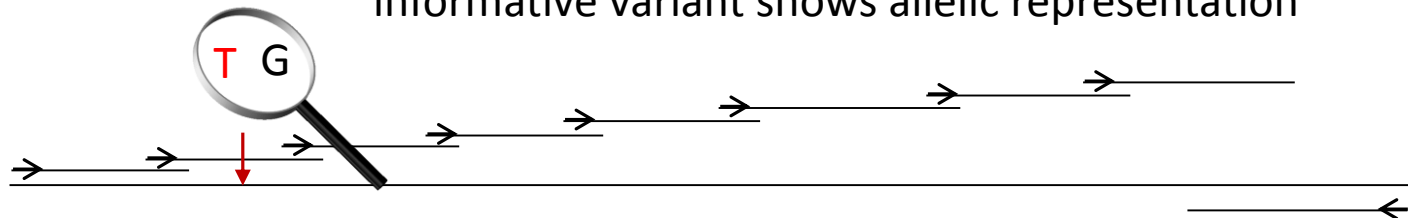
*MSH6* ~ 4.2 kb

*PMS2* ~ 2.8 kb

successful FLT amplification 98% PBL cDNA

- Sanger sequencing

complete sequence analysis in overlapping reads of one orientation  
informative variant shows allelic representation

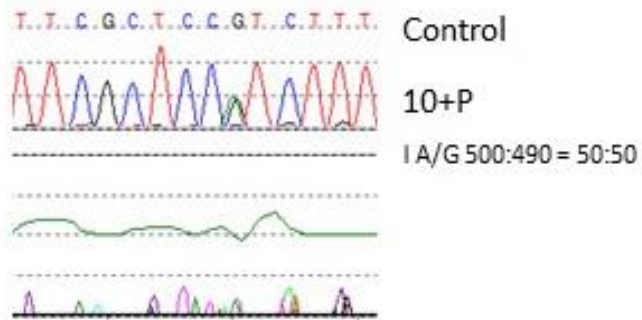




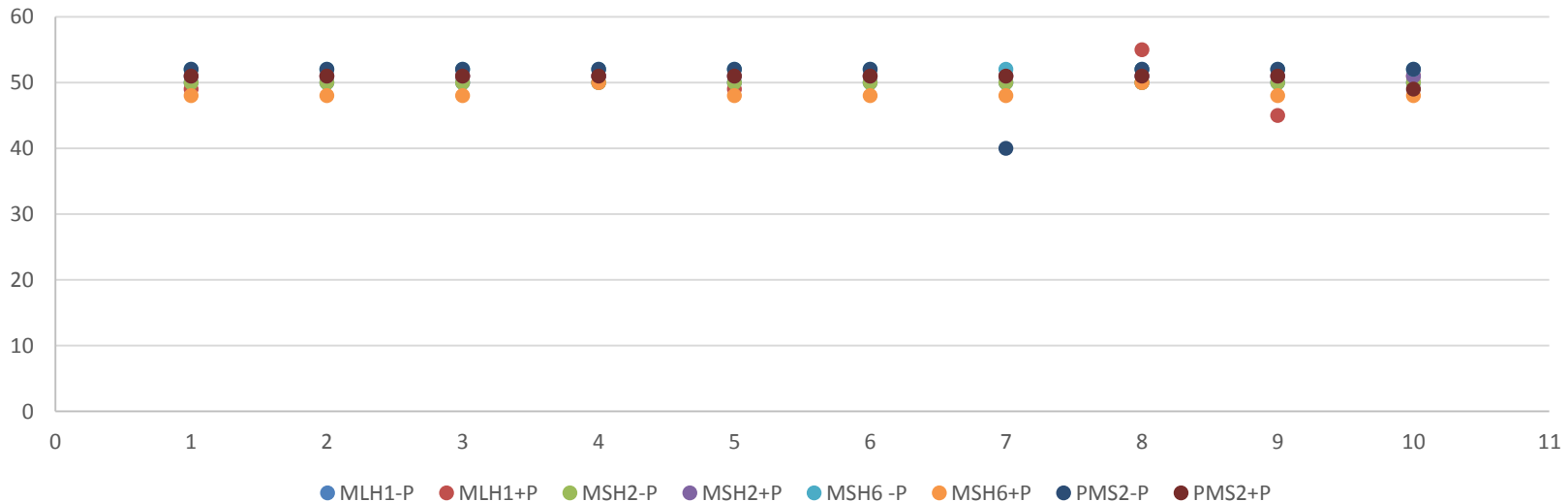
# Allelic representation in controls

peak heights in sequence electropherogram -> relative intensities in %

results compared between cDNA-P and cDNA+P



## Allelic representation MMR genes

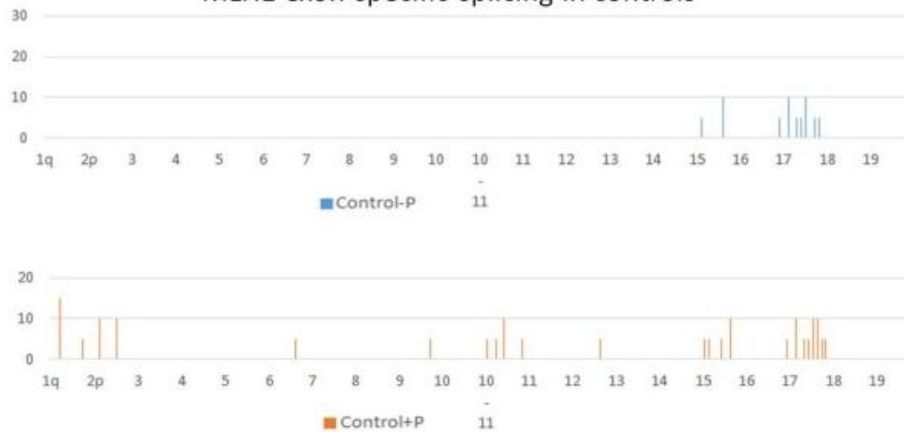




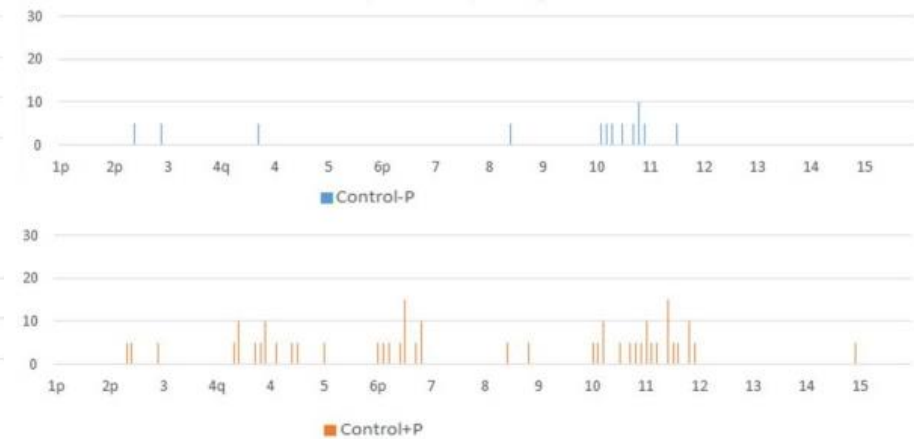
# Alternative splicing in controls

10 MMR-proficient controls for each MMR full-length transcript in cDNA -P and +P

MLH1 exon-specific splicing in controls



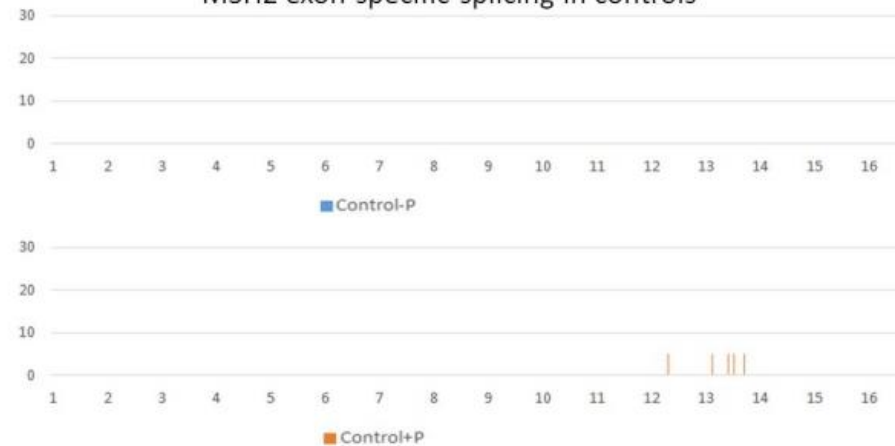
PMS2 exon-specific splicing in controls



cDNA-P

cDNA+P

MSH2 exon-specific splicing in controls



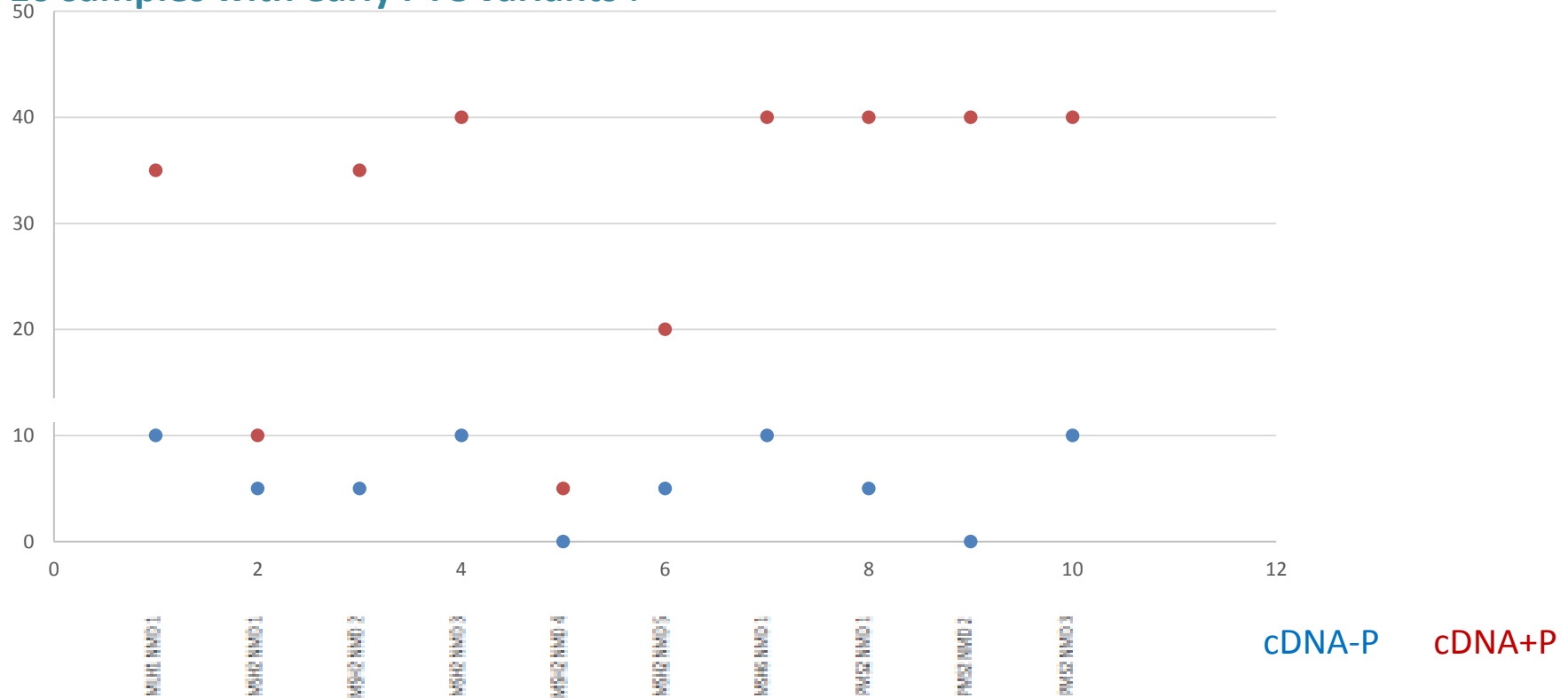
MSH6 exon-specific splicing in controls





# NMD-inhibition in controls

10 samples with early PTC variants :



-> NMD = allelic loss = AR of 0-10%

-> NMD-block usually works, but informative variant is needed for calculation of AR

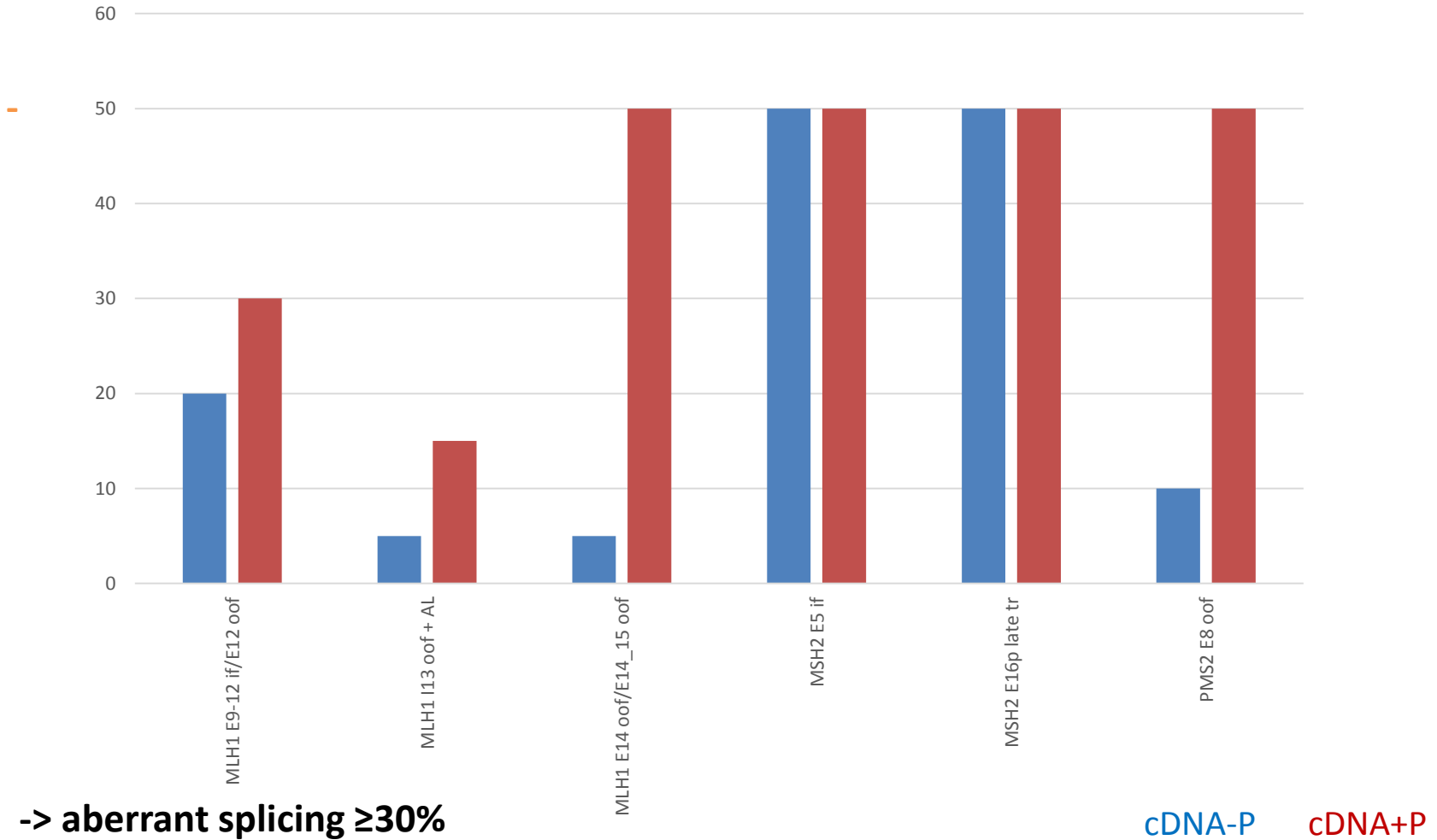
-> splicing in PTC transcripts may be enhanced (15% in 1/10 cDNA+P samples)





# Definition of aberrant splicing

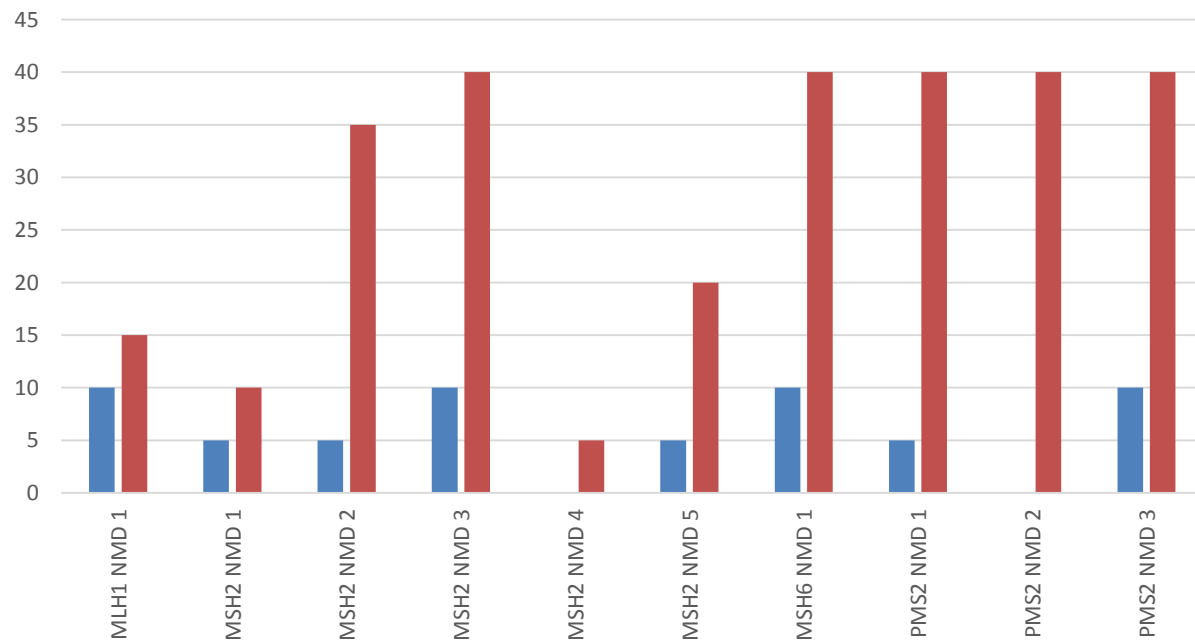
6 variants of class 4-5 predicted as splice site defect/exon skipping:



-> aberrant splicing  $\geq 30\%$

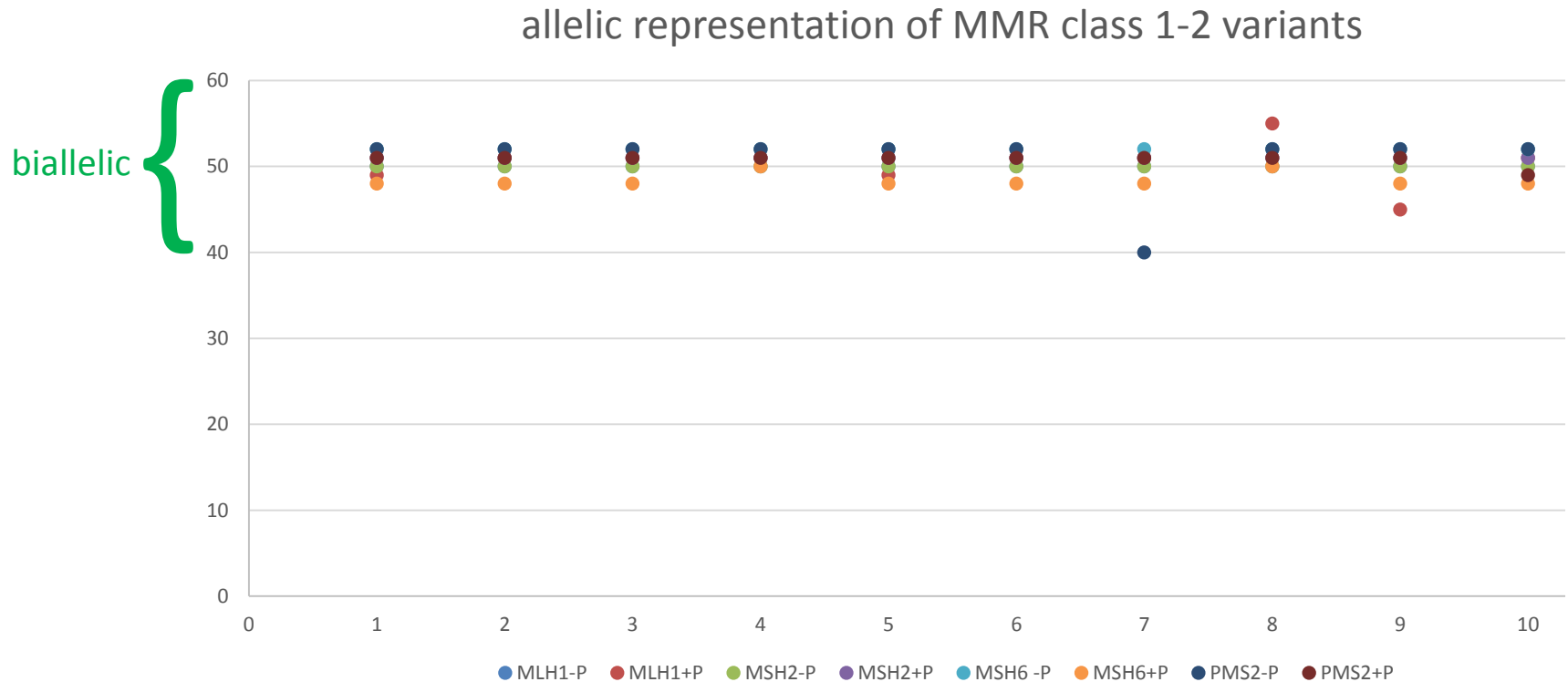
-> Splicing above alternative pattern/10%  $< 30\%$  = unclear, needs further investigation or AR

### allelic representation with active/blocked NMD



# Allelic representation of benign variants in controls

10 MMR-proficient controls, informative class 1-2 variant for each MMR gene



-> biallelic= allelic representation of 50% +/-10%

# Definition of aberrant splicing

## 6 variants of class 4-5 predicted as splice site defect/exon skipping:

### in-frame SSD

- 50±10% intensity in cDNA-P/+P

### out-of-frame SSD -> NMD

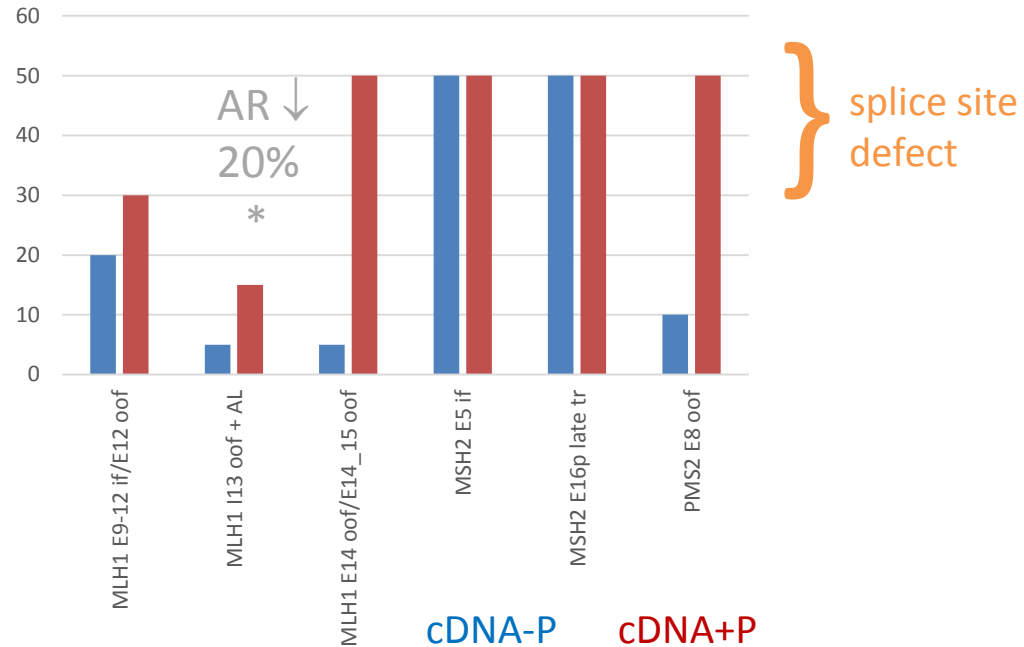
- 0-20% intensity in cDNA-P (allelic loss)
- 30-40% intensity in cDNA+P
- **exception: 15% SSD + allelic reduction 20%**

-> **aberrant splicing** ≥30%

-> Splicing above alternative pattern/10% <30% = unclear, needs further investigation or AR

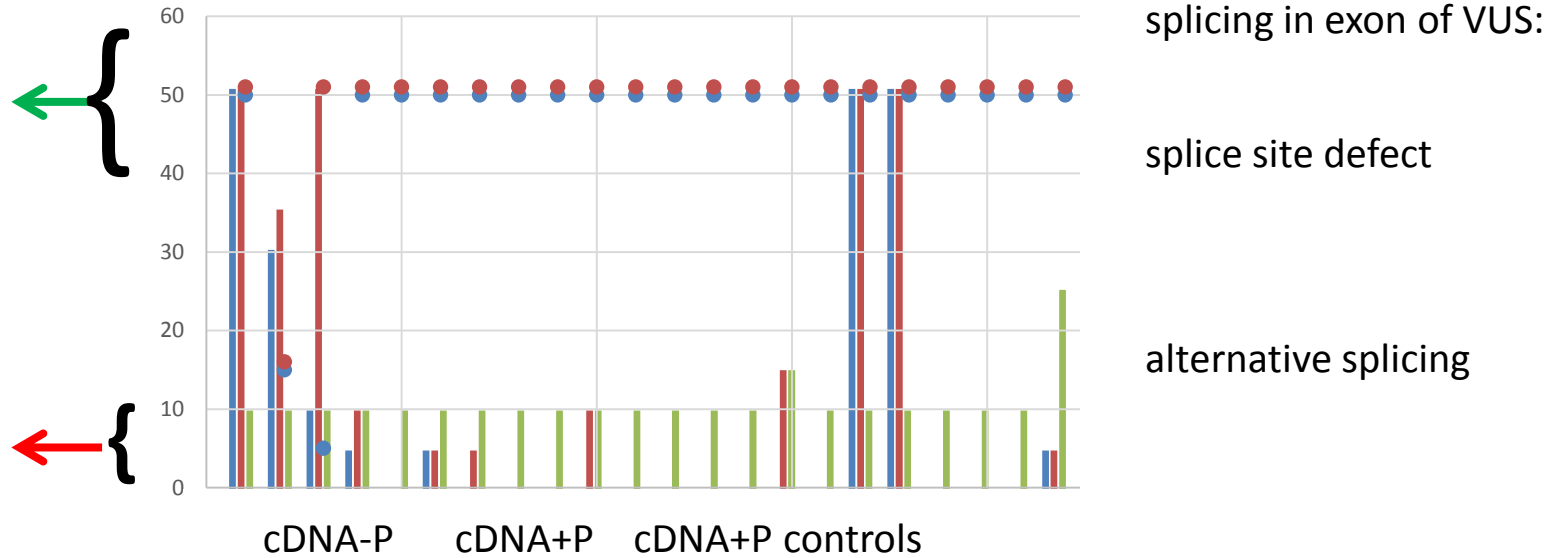
**InSiGHT: demonstrate allele-specific splicing defect in absence of wildtype transcript**

**partial splicing defects = unclear clinical relevance, class 3**



# Testing VUs

Patient samples with 25 different types of class 3 MMR variants:

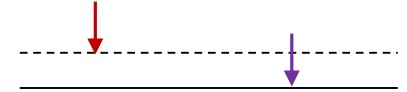


# Pros and cons of full-length transcript analysis

- universal protocol for coding MMR variants to investigate their effect on splicing

allows re-classification of VUS

determines the allelic status *in cis/trans* of VUS to a pathogenic variant

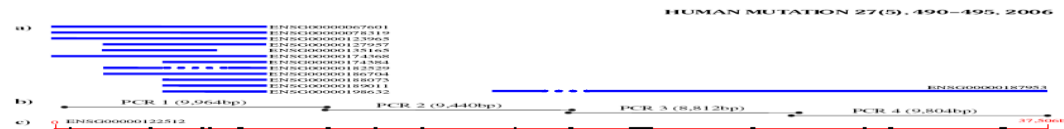


- selection for wildtype FLT

allows detection of allelic loss by informative variant (transcript integrity test)

excludes irrelevant transcripts using alternative first exons/poly-adenylation

- discrimination against *PMS2* pseudogenes



- needs informative variant for significant cDNA result (not in all samples, especially *MSH2*)

- semi-quantitative assesment of allelic representation and splicing

- requires good RNA quality for FLT amplification

- labour-intensive analysis with hands-on-time